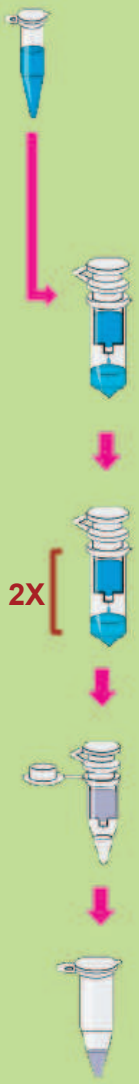
Quick Guide

MYgen Gel & PCR Purification System - Agarose Gel Extraction

Products	Cat No.	Size
MYgen Gel & PCR Purification System	MYG104-100	100 preps
	MYG104-200	200 preps

Kit Contents:
UB
WB Bottle
Help B Buffer
EB DNA Hydration Solution
Spin column / Collection tube



1. Gel block preparation in 1.5 mL tube:
   1 volume of gel cut out (100 mg)
   Add 3 volumes of **UB** buffer (300 uL)
   Incubate (65°C, 10 mins)
   Add **Isopropanol** (100 uL)
   Mix well
2. Place spin column in 2 mL collection tube
   Add 200 uL **Help B** buffer to spin column
   Centrifuge (10,000 rpm, 30 sec)
   Discard flow through
3. Transfer **Step 1** mixture into spin column from **Step 2**
   Centrifuge (7,000 rpm, 1 min)
   Discard flow through
4. Add 750 uL **WB** into spin column
   Centrifuge (13,000 rpm, 30 sec)
   Discard flow-through
   Repeat WB washing step again
5. Centrifuge (13,000 rpm, 3 mins)
6. Place spin column into new 1.5 mL tube
   Discard used 2 mL collection tube
7. Add 30-50 uL Buffer **EB**
   Incubate mixture (RT, 1 min)
   Centrifuge (13,000 rpm, 2 min)

   DNA Elution
   Store at 4°C or -20°C

Preparation:
- Prepare **80% Ethanol** (not provided) in the **WB** bottle.
- Prepare **Isopropanol** (not provided)

Note: Make fresh before use.

Protocol:

1. Weigh a new empty 1.5 mL tube. Then excise or cut out the selected DNA fragment of your choice from the agarose gel
   - Transfer the excised DNA gel fragment to the weighed 1.5 mL tube and weigh the gel fragment.
   - Add 3 volumes of **UB** buffer to 1 volume of gel
     **Eg. Add 300 uL of UB buffer to 100 mg of gel fragment*
   - Incubate the mixture for 10 mins at 65°C
   - Invert the mixture during incubation to fully dissolve gel
   - Add 1 volume of **Isopropanol** and mix well
     **Eg. Add 100 uL of Isopropanol to 100 mg of gel fragment*

   Note: For >2% gel, kindly use 6 volumes of UB buffer

2. Place the spin column in the 2 mL collection tube provided
   - Add 200 uL of **Help B** buffer to spin column
   - Centrifugation at 10,000 rpm for 30 seconds
   - Discard flow-through, place back the spin column into the 2 mL collection tube

3. Transfer mixture from **Step 1** into the spin column with 2 mL collection tube from **Step 2**
   - Centrifugation at 7,000 rpm for 1 minute
   - Discard flow-through, place back the spin column into the 2 mL collection tube

   ** To obtain high yield DNA recovery, add flow-through back in spin column and repeat step 3*

4. Add 750 uL **WB** (80% Ethanol) to the spin column
   - Centrifuge at 13,000 rpm for 30 sec
   - Discard flow-through, place back the spin column into 2 mL collection tube
   - Repeat Step 4 washing again

5. Centrifuge at 13,000 rpm for 3 mins to remove leftover residues

6. Place the spin column in a new 1.5 mL micro tube (not provided)
   - Discard the used 2 mL collection tube

7. Add 30 to 50 uL buffer **EB** DNA Hydration Solution
   - Incubate the mixture for 1 min at RT
   - Centrifuge at 13,000 rpm for 2 minute
   - Discard the spin column
   - Store at 4°C or -20°C

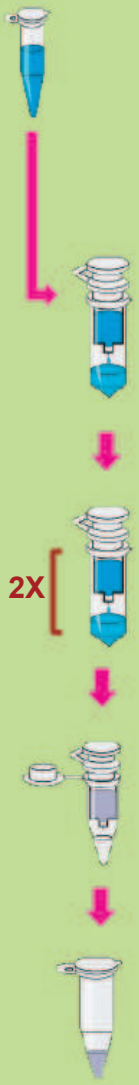
For more information or assistance, you can contact: info@gene-xpress.com.my +603-5740 8161 www.gene-xpress.com.my

Quick Guide

MYgen Gel & PCR Purification System - PCR Clean Up

Products	Cat No.	Size
MYgen Gel & PCR Purification System	MYG104-100	100 preps
	MYG104-200	200 preps

Kit Contents:
UB
WB Bottle
Help B Buffer
EB DNA Hydration Solution
Spin column / Collection tube



1. Use 1 volume of PCR sample (50 uL)
   Add 3 volumes **UB** (150 uL)
   Add 2 volumes **Isopropanol** (100 uL)
2. Place spin column in 2 mL collection tube
   Add 200 uL **Help B** buffer to spin column
   Centrifuge (10,000 rpm, 30 sec)
   Discard flow through
3. Transfer **Step 1** mixture into spin column from **Step 2**
   Centrifuge (7,000 rpm, 1 min)
   Discard flow through
4. Add 750 uL **WB** into spin column
   Centrifuge (13,000 rpm, 30 sec)
   Discard flow-through
   Repeat WB washing step again
5. Centrifuge (13,000 rpm, 3 mins)
6. Place spin column into new 1.5 mL tube
   Discard used 2 mL collection tube
7. Add 30-50 uL Buffer **EB**
   Incubate mixture (RT, 1 min)
   Centrifuge (13,000 rpm, 2 min)

   DNA Elution
   Store at 4°C or -20°C

Preparation:
- Prepare **80% Ethanol** (not provided) in the **WB** bottle.
- Prepare **Isopropanol** (not provided)

Note: Make fresh before use.

Sample Preparation: For high recovery yield of PCR product
- Use 1 volume of PCR sample (50 uL)
- Add 3 volumes of **UB** buffer (150 uL)
- Add 2 volumes of **Isopropanol** (100 uL)

Protocol:
1. Mix PCR samples as mentioned in Sample Preparation step above in a new 1.5 mL tube (not provided)

2. Place the spin column in the 2 mL collection tube provided
   - Add 200 uL of **Help B** buffer to spin column
   - Centrifugation at 10,000 rpm for 30 seconds
   - Discard flow-through, place back the spin column into the 2 mL collection tube

3. Transfer mixture from **Step 1** into the spin column with 2 mL collection tube from **Step 2**
   - Centrifugation at 7,000 rpm for 1 minute
   - Discard flow-through, place back the spin column into the 2 mL collection tube

   ** To obtain high yield DNA recovery, add flow-through back in spin column and repeat step 3*

4. Add 750 uL **WB** (80% Ethanol) to the spin column
   - Centrifuge at 13,000 rpm for 30 seconds
   - Discard flow-through, place back the spin column into 2 mL collection tube
   - Repeat Step 4 washing again

5. Centrifuge at 13,000 rpm for 3 minutes to remove leftover residues

6. Place the spin column in a new 1.5 mL micro tube (not provided)
   - Discard the used 2 mL collection tube

7. Add 30 to 50 uL buffer **EB** DNA Hydration Solution
   - Incubate the mixture for 1 minute at room temperature
   - Centrifuge at 13,000 rpm for 2 minute
   - Discard the spin column
   - Store at 4°C or -20°C

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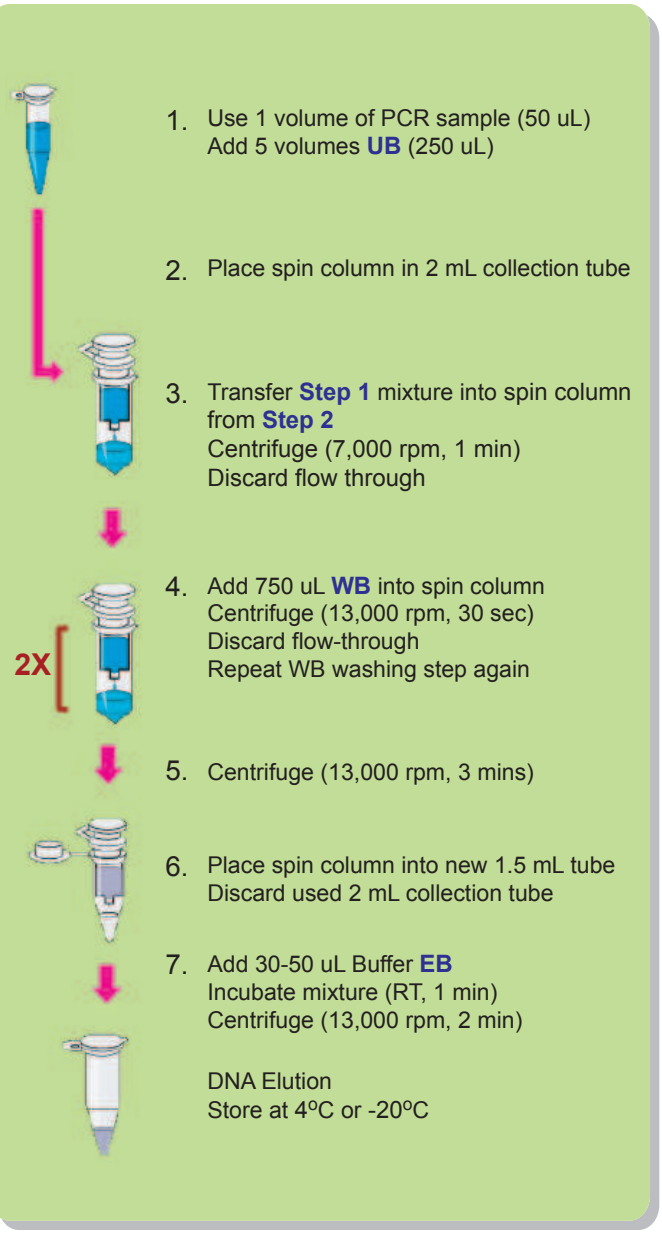
Quick Guide

MYgen Gel & PCR Purification System - Primer Dimer Removal

Products	Cat No.	Size
MYgen Gel & PCR Purification System	MYG104-100	100 preps
	MYG104-200	200 preps

Kit Contents:
UB
WB Bottle
Help B Buffer
EB DNA Hydration Solution
Spin column / Collection tube



Preparation:
- Prepare **80% Ethanol** (not provided) in the **WB** bottle.

Note: Make fresh before use.

Sample Preparation: For dimer removal from PCR product
- Use 1 volume of PCR sample (50 uL)
- Add 5 volumes of **UB** buffer (250 uL)

Protocol:
1. Mix PCR samples as mentioned in Sample Preparation step above in a new 1.5 mL tube (not provided)

2. Place the spin column in the 2 mL collection tube provided

3. Transfer mixture from **Step 1** into the spin column with 2 mL collection tube from **Step 2**
   - Centrifugation at 7,000 rpm for 1 minute
   - Discard flow-through, place back the spin column into the 2 mL collection tube

   ** To obtain high yield DNA recovery, add flow-through back in spin column and repeat step 3*

4. Add 750 uL **WB** (80% Ethanol) to the spin column
   - Centrifuge at 13,000 rpm for 30 seconds
   - Discard flow-through, place back the spin column into 2 mL collection tube
   - Repeat Step 4 washing again

5. Centrifuge at 13,000 rpm for 3 minutes to remove leftover residues

6. Place the spin column in a new 1.5 mL micro tube (not provided)
   - Discard the used 2 mL collection tube

7. Add 30 to 50 uL buffer **EB** DNA Hydration Solution
   - Incubate the mixture for 1 minute at room temperature
   - Centrifuge at 13,000 rpm for 2 minute
   - Discard the spin column
   - Store at 4°C or -20°C

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